Polymer 50 (2009) 4552-4563

Contents lists available at ScienceDirect

Polymer

journal homepage: www.elsevier.com/locate/polymer

Extrusion and characterization of functionalized cellulose whiskers reinforced polyethylene nanocomposites

Aparecido Junior de Menezes^{a,1}, Gilberto Siqueira^a, Antonio A.S. Curvelo^b, Alain Dufresne^{a,*,2}

^a Grenoble Institute of Technology, The International School of Paper, Print Media and Biomaterials (PAGORA), BP65, 38402 Saint Martin d'Hères cedex, France ^b Instituto de Química de São Carlos (IQSC), Universidade de São Paulo (USP), C.P. 780, 13560-970 São Carlos, Brazil

ARTICLE INFO

Article history: Received 31 March 2009 Received in revised form 16 July 2009 Accepted 22 July 2009 Available online 28 July 2009

Keywords: Cellulose Whiskers Nanocomposites

ABSTRACT

The surface of ramie cellulose whiskers has been chemically modified by grafting organic acid chlorides presenting different lengths of the aliphatic chain by an esterification reaction. The occurrence of the chemical modification was evaluated by FTIR and X-ray photoelectron spectroscopies, elemental analysis and contact angle measurements. The crystallinity of the particles was not altered by the chain grafting, but it was shown that covalently grafted chains were able to crystallize at the cellulose surface when using C18. Both unmodified and functionalized nanoparticles were extruded with low density polyethylene to prepare nanocomposite materials. The homogeneity of the ensuing nanocomposites was found to increase with the length of the grafted chains. The thermomechanical properties of processed nanocomposites were studied by differential scanning calorimetry (DSC), dynamical mechanical analysis (DMA) and tensile tests. A significant improvement in terms of elongation at break was observed when sufficiently long chains were grafted on the surface of the nanoparticles. It was ascribed to improved dispersion of the nanoparticles within the LDPE matrix.

© 2009 Elsevier Ltd. All rights reserved.

1. Introduction

Over the last two decades a good deal of work has been dedicated to the use of lignocellulosic fibers as reinforcing elements in polymeric matrix and for the possibility of replacing conventional fibers such as glass by natural fibers in reinforced composites [1]. However, one of the main drawbacks of lignocellulosic fibers, among others, is the important variation of properties inherent to any natural product. Indeed, their properties are related to climatic conditions, maturity, and type of soil. Disturbances during plant growth also affect the plant structure and are responsible for the enormous scatter of mechanical plant fiber properties.

One of the basic idea to achieve further improved fiber and composite is to eliminate the macroscopic flaws by disintegrating the natural grown fibers, and separating the almost defect free highly crystalline fibrils. Indeed, natural fibers display a hierarchical structure and present a multi-level organization. In fact, cellulose chains are biosynthesized by enzymes and aggregate to form microfibrils. Depending on their origin, the microfibril section ranges between 2 and 20 nm for lengths that can reach several tens of microns. They aggregate further to form fibers. Therefore, each natural fiber can be considered as a string of cellulose crystallites, linked along the chain axis by disordered domains.

Aqueous suspensions of cellulose nanocrystals can be prepared by acid hydrolysis of the biomass. The object of this treatment is to dissolve away regions of low lateral order so that the water-insoluble, highly crystalline residue may be converted into a stable suspensoid by subsequent vigorous mechanical shearing action. The resulting nanocrystals occur as rod-like particles or whiskers, which dimensions depend on the nature of the substrate, but range in the nanometer scale. Because these whiskers contain only a small number of defects, their axial Young's modulus is close to the one derived from theoretical chemistry and potentially stronger than steel and similar to Kevlar. It has been first experimentally determined in 1962 and a value of 137 GPa was reported [2]. This value differs from the theoretical estimate of 167.5 GPa reported by Tashiro and Kobayashi [3]. More recently, Raman spectroscopy technique has been used to measure the elastic modulus of native cellulose crystals. A value around 143 GPa has been reported [4]. These highly stiff nanoparticles are therefore suitable for the processing of green nanocomposite materials.

Then, the main problem is related to the homogeneous dispersion of these nanoparticles within a polymeric matrix. Because of the





^{*} Corresponding author. Tel.: +33 4 76 82 69 95; fax: +33 4 76 82 69 33. *E-mail address*: alain.dufresne@grenoble-inp.fr (A. Dufresne).

¹ Universidade Federal de São Carlos (UFSCar), 18052-780, Sorocaba, Brazil.

² Universidade Federal de Rio de Janeiro (UFRJ), Departamento de Engenharia Metalúrgica e de Materiais, Coppe, Rio de Janeiro, Brazil.

^{0032-3861/\$ -} see front matter \odot 2009 Elsevier Ltd. All rights reserved. doi:10.1016/j.polymer.2009.07.038

high stability of aqueous suspensions of cellulose whiskers, water is the preferred processing medium. Hydrosoluble polymers are therefore well adapted for the processing of cellulose whiskers reinforced nanocomposites [5]. Solid nanocomposite films can be obtained by mixing, casting and evaporating the aqueous polymer solution and the aqueous suspension. A first alternative consists in using an aqueous dispersed polymer, i.e. latex [6]. After mixing and casting the two aqueous suspensions, a solid nanocomposite film can be obtained by water evaporation and particles coalescence. A second alternative consists in using nonaqueous systems. It means that the nanoparticles should be dispersed in an adequate, with respect to the polymeric matrix, organic medium. For instance it is possible to coat the surface of nanoparticles with a surfactant [7]. The chemical modification of nanoparticles surface is another way to disperse these nanoparticles in organic solvents. It generally involves reactive hydroxyl groups from the surface [8]. Recently, it was also shown that cellulose whiskers could be dispersed in dimethylformamide, dimethyl sulfoxide or N-methyl pyrrolidine without additives or any surface modifications [9]. A solvent exchange procedure can also be used. Other possible processing techniques of nanocomposites are filtration of the suspension to obtain a film and then immersion in a polymer solution [10].

Although being widely used for the processing of thermoplastic polymers and composites reinforced with short fibers, very few studies have been reported concerning the processing of cellulose nanocrystals reinforced nanocomposites by melt extrusion methods. An attempt to prepare nanocomposites based on cellulose whiskers obtained from microcrystalline cellulose (MCC) and polylactic acid (PLA) by melt extrusion technique was recently reported [11]. The suspension of nanocrystals was pumped into the polymer melt during the extrusion process. An attempt to use polyvinyl alcohol (PVA) as a compatibilizer to promote the dispersion of cellulose whiskers within the PLA matrix was reported [12].

In the present work, cellulose whiskers were functionalized by an esterification reaction with organic acid chloride aliphatic chains of different sizes. The objective of this surface chemical treatment was to enhance the nonpolar nature of the grafted nanocrystals and improve their dispersibility in a hydrophobic polymeric matrix. Low density polyethylene (LDPE) was chosen as the matrix and the properties of ensuing extruded nanocomposites were analyzed as a function of the whiskers content and length of the grafted chains.

2. Materials and methods

2.1. Materials

Ramie fibers were obtained from Stucken Melchers GmbH & Co. (Germany). The low density polyethylene (LDPE) used for the study was PB-608 obtained from Braskem. It has a room temperature density of 0.915 g cm⁻³ and Melt Flow Index (190 °C/2.16 kg) of 30 g/10 min. Sulfuric acid (95%), triethylamine (TEA, 99.5%), toluene (anhydrous, 99.8%), acetone (99%), hexanoyl chloride, lauroyl chloride and stearoyl chloride (99%) were all obtained from Sigma-Aldrich. The chemical structures of the various chemical grafting agents as well as their typical dimensions compared to cellulose whiskers are reported in Fig. 1.

2.2. Preparation of cellulose nanocrystals or whiskers

Ramie fibers were first cut in small pieces and treated with a 2% NaOH solution at 80 °C for 2 h to remove residual additives. Then, the ramie fibers were submitted to an acid hydrolysis treatment with a 65 wt% H_2SO_4 solution at 55 °C for 30 min under continuous stirring. The suspension was washed with water until neutrality and dialyzed with deionized water. The obtained suspension was homogenized with an Ultra Turrax T25 homogenizer at 13,500 rpm (2–5 min) and then filtered in sintered glass No 1.

2.3. Surface chemical modification of cellulose whiskers

The surface chemical modification of the cellulose whiskers was performed in a round-bottomed reaction flask under reflux (4 h) and under constant mechanical stirring in toluene medium. The toluene suspension was obtained by a solvent exchange procedure and centrifugation procedure (water to acetone, and then acetone to toluene, both four times). Ramie nanocrystals (2 g) were mixed with triethylamine (5 mL) and organic acid chloride (5.2 mL for hexanoyl, 8.8 mL for lauroyl or 12.5 mL for stearoyl chloride). Triethylamine was utilized to catalyze the reaction and as a complexing agent for HCl formed during the reaction [13]. The modified nanocrystals were submitted to a Soxhlet extraction with acetone for 24 h. The ramie cellulose whiskers modified with hexanoyl chloride, lauroyl chloride and stearoyl chloride will be denoted WRC6, WRC12 and WRC18,



Fig. 1. Chemical structures of the various chemical grafting agents and their typical dimensions compared to cellulose whiskers.

respectively, whereas unmodified whiskers will be later on denoted WRU. The scheme of the reaction was published elsewhere [13].

2.4. Processing of nanocomposite materials

Nanocomposite materials were prepared by mixing LDPE and either unmodified or chemically modified ramie cellulose whiskers (whiskers content ranging from 0 to 15 wt%) using a twin-screw DSM Micro 15 compounder. The filler was used in the dry state after water or toluene evaporation for unmodified and functionalized cellulose whiskers, respectively. The components were introduced in the mixing chamber and allowed to melt at 160 °C. The mixing speed was set at 60 rpm for 10 min. The ensuing mixture was hotpressed using a laboratory press (St-Eloi Mécanique) at 160 °C under a force of 10 ton. This temperature was low enough, even under shear during the extrusion process, to avoid cellulose degradation.

2.5. Characterization

2.5.1. FTIR spectroscopy

FTIR spectrograms were obtained on a Perkin–Elmer Paragon 1000 FTIR spectrometer. Both unmodified and chemically modified



Fig. 2. Transmission electron micrograph (A) and length and diameter histograms and cumulative curves of ramie cellulose whiskers (B).



Fig. 3. FTIR spectra (KBr pellets) (A) and general XPS spectra (B) of unmodified ramie cellulose whiskers (a) and surface modified with hexanoyl chloride (b), lauroyl chloride (c) and stearoyl chloride (d), after Soxhlet extraction with acetone.

ramie cellulose whiskers were analyzed as KBr pellets (1:100). The spectra were obtained with a resolution of 1 cm^{-1} , averaging over 16 scans.

2.5.2. XPS spectroscopy

X-ray photoelectron spectrograms were performed using a XR3E2 (Vacuum Generator, UK) instrument equipped with monochromated Mg K α X-ray source (1253.6 eV) and operated at 15 kV under a current of 20 mA.

2.5.3. Elemental analysis

Duplicate elemental analysis was carried out at the Laboratoire Central d'Analyses de Vernaison, France (CNRS). This technique is based on atomic absorption of the investigated elements. The carbon, nitrogen, and oxygen contents of cellulose nanocrystals were measured independently.

2.5.4. X-ray diffraction

X-ray diffraction data were recorded in reflection mode for dry whiskers powder and LDPE based nanocomposite films at room temperature with a Siemens D500 diffractometer equipped with a CuK α anode (λ = 0.15406 nm).

2.5.5. Transmission electron microscopy (TEM)

Drops of cellulose whiskers suspension were deposited on carbon-coated electron microscope grids and negatively stained with uranyl acetate solution. A Philips transmission electron microscope at an acceleration voltage of 80 kV was used.

2.5.6. Contact angle measurements

Contact angle measurements were performed at room temperature with an OCA20 (DataPhysiscs Instruments). Three different liquids (water, *N*,*N*-dimethyl formamide and diiodomethane) with different dispersive and polar surface tensions were used to determine the surface energy of both unmodified and modified ramie whiskers, using the approach proposed by Owens and Wendt [14].

2.5.7. Thermogravimetric analysis (TGA)

TGA was carried out with a Shimadzu TA-50 instrument at 20 °C min⁻¹ in a nitrogen atmosphere, from 25 to 900 °C. Samples were obtained by cutting the plates in circular form and weighed.

2.5.8. Differential scanning calorimetry (DSC)

DSC was performed using a DSC Q100 differential scanning calorimeter from TA Instruments. Samples were heated from -100 to $150 \,^{\circ}C$ at a heating rate of $10 \,^{\circ}C \,min^{-1}$ under N_2 atmosphere.

2.5.9. Dynamic mechanical analysis (DMA)

DMA measurements were performed with a RSA3 (TA Instruments) working in the tensile mode. The sample dimensions were $10 \times 5 \times 0.2 \text{ mm}^3$ and tests were performed under isochronal conditions at 1 Hz and the temperature was varied between -100 and $110 \,^{\circ}\text{C}$ at a heating rate of 5 $^{\circ}\text{C}$ min⁻¹.

2.5.10. Tensile tests

The nonlinear mechanical properties of the composites were carried out with an RSA3 (TA Instruments) with a load cell of 100 N. Experiments were performed at room temperature with a crosshead speed of 10 mm min⁻¹. The sample dimensions were $10 \times 5 \times 0.2 \text{ mm}^3$ and the results were averaged over five measurements.

3. Results and discussion

3.1. Characterization of ramie cellulose whiskers

3.1.1. Morphological analysis

Acid hydrolysis of native ramie cellulose fibers leads to aqueous suspensions of elongated nanocrystals with high aspect ratio. Fig. 2A shows an electron micrograph of ramie cellulose whiskers. The length and diameter of these nanocrystals were determined by using digital image analysis (ImageJ). Both the obtained histograms and cumulative curves are shown in Fig. 2B. The geometric average length and diameter were around 134 nm \pm 59 nm and 10.8 nm \pm 4.5 nm, respectively, giving rise to an aspect ratio around 12. A minimum of 228 and 70 measurements were used to determine the length and the diameter, respectively, of ramie whiskers. It is worth noting that more than 50% of the nanoparticles have a length lower than 100 nm.

Regarding their arrangement in solution, it is noteworthy that because of electrostatic repulsions between surface-grafted sulfate ester groups resulting from the sulfuric acid hydrolysis, the cellulose whiskers repel each other and then do not flocculate in water.

Table 1

XPS analysis of ramie cellulose whiskers after and before surface chemical modification with hexanoyl, lauroyl and stearoyl chloride.

Sample	0/C	Binding energy (eV)				
		C1, 285 \pm 0.1, C-C/C-H	C2, 286.4 \pm 0.1, C–O	C3, 287.7 \pm 0.1, 0-C-0/C=0	C4, 289 ± 0.1 , 0-C=0	(%)
WRU	0.51	27.4	52.8	17.2	2.6	-
WRC6	0.53	34.5	46.4	14.6	4.5	7.1
WRC12	0.41	45.0	38.4	12.6	3.9	17.6
WRC18	0.36	50.4	37.8	9.8	1.9	23.0



Fig. 4. Decomposition of C1s signal into its constituent contributions for unmodified ramie cellulose whiskers (A) and modified with hexanoyl chloride (B), lauroyl chloride (C) and stearoyl chloride (D).

The overall concentration of these sulfate moieties is related to the sulfur ratio, which was determined by elementary analysis to be 0.57% of dry matter [15]. According to these sulfur ratios and to the average geometry of cellulose nanocrystals, the average surface charge of rods was estimated to 0.60 e nm⁻² (considering rod-like nanoparticles with an average diameter of 6–8 nm and a length of about 150–250 nm).

3.1.2. FTIR investigation

The FTIR spectra recorded for both unmodified and chemically modified ramie whiskers are shown in Fig. 3A. Compared to unmodified whiskers, the spectra corresponding to modified cellulose nanoparticles display an extra peak at 1737 cm⁻¹, attributed to

carbonyl groups. The signals at 2953, 2919 and 2850 cm^{-1} are ascribed to the presence of grafted alkane chains. The concomitant decrease of the magnitude of the broad band around 3300 cm^{-1} for modified whiskers compared to unmodified is attributed to the partial disappearance of OH groups, confirming the success of the grafting reaction with organic acid chlorides.

3.1.3. XPS analysis

Fig. 3B shows the XPS spectra for both unmodified and chemically grafted ramie whiskers. The signals observed around binding energies of 531 and 287 eV correspond to the 1s orbital electron of oxygen and carbon, respectively. The elemental surface composition (%) and the oxygen-to-carbon ratios of the different samples are



Fig. 5. Evolution of the surface coverage area and degree of substitution (DS) as a function of the length of grafted chains. The lines serve to guide the eyes.

 Table 2

 Elemental analyses of ramie cellulose nanocrystals before and after chemical modification with hexanoyl, lauroyl and stearoyl chloride.

Sample	C (%)	H (%)	O (%)	DS
WRU	43.08	6.41	50.51	-
WRC6	52.90	7.67	39.43	0.68
WRC12	53.57	8.12	38.31	0.32
WRC18	56.90	8.88	34.22	0.31

summarized in Table 1. It decreases from 0.51 for the unmodified cellulose whiskers to 0.36 for the sample grafted with C18 chains. It is also observed that the O/C ratio decreases with increasing the length of the aliphatic grafted chains typical of organic acid chloride aliphatic chains. Fig. 4 presents the deconvolution of the C1s peak that is composed of the C1 peak corresponding to C–C/C–H linkages, C2 peak corresponding to C–O of alcohols and ethers, C3 peak corresponding to the O–C–O and C==O from the acetal moieties and C4 peak, corresponding to O–C==O and representing the ester carbon contribution. The peak corresponding to carbon was deconvoluted for each sample using curve fitting software (Spectrum NT).

Considerable changes can be observed in the intensities of the C1 peak, referring to aliphatic chains (C–H), for modified samples when compared to the pristine sample. This behavior can be attributed to the chain grafting at the whisker surface and provides additional proof of successful grafting of aliphatic chains to cellulose. Indeed, XPS is a powerful tool to investigate chemical changes resulting from surface modification since the investigated thickness is of the order of about 6 nm. Values of the C1, C2, C3 and C4 contributions are reported in Table 1. The surface coverage by large alkane chain (θ_{SC}) is confirmed, as estimated by the C1 relative area according to equation (1) [16].

$$\theta_{\rm SC} = A_{\rm C1m} - A_{\rm C1um} \tag{1}$$

where, A_{C1m} and A_{C1um} are the areas of the modified and unmodified samples, respectively. The θ_{SC} values are reported in Table 1 and its evolution as a function of the number of carbons of the grafted chain is shown in Fig. 5. The increase of the surface coverage



Fig. 6. Wide-angle X-ray diffraction patterns for ramie cellulose whiskers (A): unmodified (a) and modified with hexanoyl chloride (b), lauroyl chloride (c) and stearoyl chloride (d), and nanocomposite films (B): neat LDPE matrix (a) and related nanocomposite films reinforced with 10 wt% ramie cellulose whiskers: unmodified (b) and modified with hexanoyl chloride (c), lauroyl chloride (d) and stearoyl chloride (e).

Table 3

Contact angles and surface tension contributions of the ramie cellulose whiskers before and after modification with hexanoyl, lauroyl and stearoyl chloride.

Sample	Contact angle			$\gamma \xi(mJ/m^2)$	$\gamma^d_S(mJ/m^2)$	$\gamma_{\rm S}^{\rm T} ({\rm mJ}/{\rm m}^2)$
	Water	Formamide	Diiodomethane			
WRU	35	19	40	35.2	25.3	60.5
WRC6	72	40	61	12.5	25.6	38.1
WRC12	90	55	61	2.9	30.0	32.9
WRC18	101	75	56	0.2	31.3	31.5

gives a clear evidence of the occurrence of the expected surface chemical modification of the ramie cellulose whiskers.

3.1.4. Elemental analysis

The elemental analyses of both unmodified and modified samples are given in Table 2. The degree of substitution (DS) of the modified ramie whiskers was determined from the data obtained by elemental analysis [17]. The DS values obtained from this technique are reported in Table 2 and its evolution as a function of the number of carbons of the grafted chain is shown in Fig. 5. A decrease of DS is observed when the number of carbon atoms of the chloride acid increases and it seems to stabilize for highest chain lengths. This behavior was also observed after esterification of lignocellulosic materials [18].

3.1.5. X-ray diffraction

Natural lignocellulosic fibers are known to display X ray diffraction (XRD) patterns typical of cellulose type I, with the main diffraction signals at 2θ values of 15° , 16° , $22,5^{\circ}$ and 34° , attributed to the diffraction planes 101, 101, 002 and 040, respectively. Fig. 6A shows the XRD patterns obtained for the modified ramie cellulose whiskers samples as well as the one corresponding to the pristine sample. Even after chemical modification, the cellulosic nanoparticles remain semicrystalline and display the same XRD patterns leading to the conclusion that the initial crystallinity was retained. Then, the surface chemical modification did not alter the crystallinity of cellulose nanocrystals. Upon chemical modification, a new ill-defined peak, located by an arrow, appears around 21° that is attributed to the presence of grafted aliphatic chains. In a previous work [19], a similar but much more defined peak was observed for polycaprolactone-grafted starch nanocrystals. This peak was much less defined for PCL-grafted cellulose whiskers because a strong



Fig. 7. Water contact angle with time for unmodified ramie cellulose whiskers (a) and modified with hexanoyl chloride (b), lauroyl chloride (c) and stearoyl chloride (d).



Fig. 8. TGA curves (A) and dTGA (B) for unmodified ramie cellulose whiskers (a) and modified with hexanoyl chloride (b), lauroyl chloride (c) and stearoyl chloride.

diffraction peak due to cellulose I occurred in the same diffraction angle range.

3.1.6. Contact angle measurements

Table 3 shows the values of contact angle measured for ramie cellulose whiskers at longer times, i.e. when the equilibrium was reached. The values of the surface energy as well as the polar and dispersive contributions are also reported. The dynamic behavior of the contact angle for a drop of distilled water onto the surface of the materials is shown in Fig. 7. The untreated cellulose surface gave the lowest initial contact angle value, which is an expected finding since the surface of cellulose nanoparticles contains OH-rich macromolecules, the most capable to establish hydrogen bonds with water. The initial contact angle is two times higher for grafted substrates, indicating that chemical treatments induced dramatic changes in surface polarity of cellulose nanocrystals. The contact angle value increases as the length of the grafted chain increases. Moreover, whereas the contact angle value decreases with time for WRU and WRC6 substrates, it remains almost constant for WRC12 and WRC18. Then, contact angles measurements give a clear evidence of the change in hydrophobicity of the modified samples compared to the unmodified sample. These results were confirmed by the decrease of the polar contribution to the surface energy (Table 3).

The initial contact angle values, corresponding to times ~ 0 , for WRU and WRC6 are 45 and 87, respectively. After few seconds, it remains almost constant with values of 35 and 72. It was checked that the decrease of the contact angle value was due a spreading of the drop rather than a penetration by capillarity because of the porous character of the pellets prepared for these measurements. It was checked that the diameter of the drop increased and that the volume of the drop remained constant during time.

3.1.7. Thermal analysis

The thermal degradation behavior of the cellulose nanocrystals was investigated from TGA measurements. Results are reported in Fig. 8. The unmodified sample displays a weight loss from room temperature to 130 °C. It is ascribed to the presence of water. This effect is decreased for the sample modified with C6 and absent for other specimens, i.e. nanocrystals modified with C12 and C18. It is obviously ascribed to a lower accessibility of surface OH groups after the grafting reaction. Then, a higher weight loss is observed in the range 250–350 °C. The thermal decomposition temperatures, associated with a 2% weight loss and maximum of derived signal, were determined and results are collected in Table 4. The degradation temperature is lower for modified nanoparticles compared

to the pristine cellulose whiskers and it decreases as the length of the grafted chains increases as already reported for another system [20].

The DSC thermograms obtained for both unmodified and chemically modified nanoparticles are shown in Fig. 9. Panel A corresponds to the first temperature scan, whereas panel B corresponds to the second temperature scan, the samples being quenched between each temperature scan. An endothermic peak is observed around 110 °C for the pristine sample during the first temperature scan. This peak is not observed during the second temperature scan and was ascribed to the vaporization of water. For modified samples, it is not observed because of the increased hydrophobicity of the nanoparticles. However, the thermogram recorded for cellulose whiskers modified with C18 during the first temperature scan (Fig. 9A-curve d) shows a double endothermic peak between 25 and 80 °C. It is ascribed to the melting of covalently linked chains at the cellulose nanocrystal surface that were sufficiently long to crystallize. The formation of a crystalline brushlike structure of stearate moieties grafted to starch nanoparticle surface was reported elsewhere [13]. Even after quenching, a melting endotherm is observed for the WRC18 sample (Fig. 9B) showing that the kinetics of crystallization of grafted chains is fast. However, the global aspect of this melting endotherm is different from the one recorded during the initial temperature scan.

3.2. Characterization of ramie cellulose whiskers reinforced LDPE nanocomposite films

3.2.1. Morphological analysis

Fig. 10 shows photographs of the unfilled LDPE film and ramie cellulose whiskers based nanocomposite films reinforced with 10 wt% of WRU and WRC18. The neat PE is obviously translucent as any low thickness polymeric film with a relatively low degree of

Table 4

Degradation data obtained from TGA measurements for unmodified and modified ramie whiskers: T_{onset} corresponds to the beginning of the degradation process, and T_{d1} and T_{d2} to the thermal decomposition temperature associated with a 2% weight loss and maximum of derived signal, respectively. Measurements were performed under nitrogen flow at 20 °C min⁻¹.

Samples	T_{onset} (≤0.5% weight loss)	T _d (°C) (at 2% weight loss)	<i>T</i> _d maximum by dTGA (°C)
WRU	210	240	368
WRC6	242	267	320
WRC12	200	236	321
WRC18	215	248	305



Fig. 9. DSC traces for unmodified ramie cellulose whiskers (a) and modified with hexanoyl chloride (b), lauroyl chloride (c) and stearoyl chloride (d). First temperature scan (A) and second temperature scan recorded after the first temperature scan and subsequent quenching (B). Traces have been shifted vertically.

crystallinity. When adding 10 wt% of cellulose whiskers, the film becomes dotted with black. These heterogeneities reveal the poor and inhomogeneous dispersion of the filler within the polymeric matrix. It is obviously ascribed to the highly hydrophobic nature of the matrix and highly hydrophilic character of the cellulose nanoparticles. When the cellulose whiskers were chemically modified with aliphatic chains, the occurrence of these aggregates progressively vanish and the appearance of the composite film reinforced with WRC18 becomes similar to the one of the unfilled film. In addition, it is worth noting that it seems that the cellulose whiskers are not orientated when extruded in the melt. Indeed, we observed that no difference was reported for extruded films reinforced with cellulose whiskers when testing the film in the flow or transverse direction. These results are reported in a forthcoming publication.

3.2.2. Thermal analysis

The thermal characterization of ramie cellulose whiskers reinforced LDPE nanocomposite films was carried out using DSC. From the analysis of DSC traces, the melting temperature (T_m), associated heat of fusion (ΔH_m) and degree of crystallinity (χ_c) were obtained for the unfilled LDPE film, and nanocomposite materials reinforced with either unmodified or modified whiskers. The resulting experimental data are listed in Table 5. It is worth noting that for the calculation of the degree of crystallinity of the composite materials, the heat of fusion was normalized to the matrix content.

The melting point remains roughly constant between 103 and 105 °C upon whiskers addition, regardless their modification state. It is an indication that the size of the crystallites is not affected by the filler. On the contrary, the degree of crystallinity of the LDPE matrix was found to increase with the whiskers content nearly independently of their nature (Fig. 11), their surface being chemically modified or not. It seems that the cellulosic nanoparticles



Fig. 10. Photographs of the neat film and ramie cellulose whiskers based nanocomposite films reinforced with 10 wt% of WRU and WRC18.

probably act as nucleating agents for the polymeric matrix. This nucleating effect is surprisingly not influenced by the grafting of aliphatic chains at the nanoparticle surface. Indeed, because it has been shown from DSC measurements that, at least for WRC18 nanoparticles, grafted chains can crystallize at the surface of cellulose whiskers, one could expect a possible co-crystallization of covalently linked chains at the nanoparticles surface with those from the matrix. However, it is worth noting that for modified whiskers reinforced composites, the nanocrystal content refers to the weight fraction of grafted nanoparticles and that the effective whisker content is lower compared to unmodified. It means that experimental data for modified nanoparticles based composites in Fig. 11 should be shifted horizontally towards the lower whiskers content.

The thermal degradation behavior of ramie cellulose whiskers reinforced LDPE nanocomposite films was investigated from TGA measurements. Results are reported in Table 6. The onset temperature of the weight loss process was systematically lower for composites than for the neat matrix. It is ascribed to the water content of the cellulosic filler. The degradation temperature associated to the maximum of the dTGA signal remains roughly constant regardless the cellulose whiskers content and its surface modification.

Table 5

Melting characteristics of LDPE-based nanocomposites reinforced with ramie cellulose whiskers obtained from DSC measurements: melting temperature (T_m), enthalpy of fusion (ΔH_m) and degree of crystallinity (χ_c).

Sample	Whisker Content (%)	$T_{\rm m}(^{\circ}{\rm C})$	$\Delta H_{\rm m}$ (J g ⁻¹)	χ_c^a
LDPE	0	103	110.1	0.38
LDPE-WRU	3	104	114.0	0.41
	5	103	113.4	0.41
	10	103	127.2	0.49
	15	103	124.0	0.51
LDPE-WRC6	3	104	110.2	0.39
	5	103	116.3	0.43
	10	104	124.3	0.48
	15	105	127.1	0.52
3LDPE-WRC12	3	103	112.4	0.40
	5	104	119.2	0.44
	10	103	130.9	0.51
	15	104	125.2	0.51
LDPE-WRC18	3	105	115.7	0.41
	5	103	122.0	0.45
	10	105	132.9	0.51
	15	104	126.1	0.52

^a $\chi_c = \Delta H_m/w\Delta H_m^\circ$, where $\Delta H_m^\circ = 290 \text{ J/g}$ (heat of fusion for 100% crystalline LDPE) and *w* is the weight fraction of polymeric matrix in the composite.



Fig. 11. Evolution of the degree of crystallinity of the LDPE matrix as a function of the ramie cellulose whiskers content for nanocomposite films reinforced with unmodified (\bullet) and modified with hexanoyl chloride (\circ), lauroyl chloride (\blacktriangle) and stearoyl chloride (\triangle) whiskers. The lines serve to guide the eyes.

3.2.3. X-ray diffraction

Fig. 6B shows the X-ray diffraction patterns obtained for the neat LDPE matrix and related nanocomposites reinforced with 10 wt% cellulose whiskers. No alteration of the diffraction pattern of LDPE is observed upon ramie whiskers addition indicating that the crystallinity of LDPE is not affected upon whiskers addition. However, it is worth noting that there might be local changes in crystallinity that the diffractometer cannot spatially resolve.

3.2.4. Mechanical properties

Fig. 12 shows the evolution of the logarithm of the storage tensile modulus as a function of temperature for ramie cellulose whiskers reinforced LDPE nanocomposites. Panels A, B, C and D correspond to filler contents of 3, 5, 10 and 15 wt%, respectively. The behavior of the neat LDPE matrix has been added in each figure for reference and modulus values have been normalized at low temperature. The modulus drop observed around -40 °C is ascribed to the anelastic manifestation of the glass transition of the polymeric matrix. The modulus drop corresponding to this relaxation is weak because LDPE is a semicrystalline polymer. Indeed, the rubbery modulus is known to depend on the degree of crystallinity of the material, the crystalline regions of LDPE acting as physical cross-links for the elastomer. In this physically cross-linked system, the crystalline regions would also act as filler particles due to their finite size, which would increase the modulus substantially. At higher temperatures, E' decreases continuously because of the progressive melting of LDPE. At the melting point of the polymeric matrix, the modulus drops irreversibly and the setup fails to measure it, due to irreversible chain flow.

When adding ramie whiskers, the rubbery modulus slightly increases. This low but significant increase can be ascribed to a reinforcing effect of the cellulose whiskers and/or to the increase of the degree of crystallinity reported from DSC measurements. Surprisingly, no effect of the chemical grafting of aliphatic chains on the surface of the nanoparticles is observed. This is probably ascribed to two antagonist effects. Indeed, the chemical grafting should improve the dispersion of the filler within the polymeric matrix, but at the same time it decreases the possibility of interwhiskers interactions. These interactions were reported to be the basis of the reinforcing effect for cellulose whiskers reinforced nanocomposites [21]. However, it is worth remember that for modified whiskers reinforced composites, the effective whisker content is lower than for its unmodified counterpart as previously emphasized.

In addition, most of the nanocomposite films display a two-steps modulus drop in the rubbery region of the LDPE matrix. Indeed, apart the one associated as for the neat matrix to the glass transition of LDPE, a high temperature modulus drop, around 35 °C is observed. This new relaxation process is well identified through the maximum of the loss angle in Fig. 13. The existence of the second $\tan \delta$ peak could suggest that the main relaxation process, associated with T_g of the polymeric matrix, splits into two well-defined peaks. This splitting of the relaxation process could be ascribed to strong interactions between the functionalized cellulosic nanoparticles and the LDPE matrix. These interactions could lead to the formation of an interfacial layer surrounding the filler and which mobility is restricted compared to the bulk matrix. This phenomenon could be obviously emphasized because of the nanometric scale of the filler and omnipresence of the surface. However, if such an effect is at the origin of the splitting of the relaxation process, then the relative magnitude of the high temperature peak should increase when increasing the whiskers content, which was not observed

The second explanation is related to the possible cellulose whiskers-induced crystallization of the polymeric matrix. This explanation is highly speculative but the origin of this modulus drop remains mysterious. It is worth noting that the temperature of this second peak corresponds to the one of the melting endotherm observed for WRC18 in Fig. 9.

Tensile tests were performed at room temperature. From the obtained stress-strain curves, the strength, tensile modulus and elongation at break for ramie cellulose whiskers reinforced LDPE were determined. Fig. 14 shows the evolution of these parameters as a function of the whiskers content. A slight increase of the tensile modulus (Fig. 14B) and a slight decrease of the tensile strength (Fig. 14A) upon whiskers addition are observed. Again, no discriminating effect of the filler-matrix compatibilization is reported. However, as for other tests the comparison is difficult because both the effective whiskers content and the degree of crystallinity of the specimen differ depending on the chemical modification of the filler.

Table 6

Thermal degradation characteristics (T_{onset} : onset of the degradation, T_d : thermal decomposition temperature associated with a 2% weight loss, and T_d maximum: maximum of the derived signal) for LDPE-based nanocomposite films reinforced with ramie cellulose whiskers immersed in water.

Sample	Whisker content (%)	$T_{\text{onset}} (\geq 0.5\%$ weight loss)	T _d (°C) (at 2% weight loss)	T _d maximum by dTGA (°C)
LDPE	0	321	354	482
LDPE-WRU	3	263	310	488
	5	255	297	487
	10	247	283	484
	15	239	268	485
LDPE-WRC6	3	293	334	480
	5	280	306	485
	10	276	293	487
	15	271	288	488
LDPE-	3	297	333	490
WRC12	5	281	320	474
	10	286	310	487
	15	281	302	493
LDPE-WRC18	3	297	335	491
	5	290	324	495
	10	275	301	489
	15	256	282	480



Fig. 12. Evolution of the logarithm of the storage tensile modulus as a function of temperature for the neat LDPE matrix () and related nanocomposite films reinforced with 3 (A), 5 (B), 10 (C) and 15 wt% ramie cellulose whiskers: unmodified () and modified with hexanoyl chloride (), lauroyl chloride () and stearoyl chloride ().

The elongation at break decreases upon whiskers addition as expected and tends to roughly stabilize at high filler loading (Fig. 14C). However, an interesting feature is observed. When the grafted chains are sufficiently long (C18-grafted nanoparticles), the elongation at break is systematically much higher for a given composition than for other samples. It could be due to a possible co-crystallization between covalently linked chains at the surface of the

nanoparticles with those from the matrix, but this co-crystallization was not evidenced from other measurements. Entanglements between grafted chains and macromolecular chains from the polymeric matrix are highly unlikely because of the low molecular weight of the grafted moieties. The most probable explanation could be the highest dispersion level of cellulose whiskers chemically modified with C18 chains.



Fig. 13. Evolution of the tangent of the loss angle (tan δ) as a function of temperature for the neat LDPE matrix (\bullet) and related nanocomposite films reinforced with 3 (A), 5 (B), 10 (C) and 15 wt% ramie cellulose whiskers: unmodified (\circ) and modified with hexanoyl chloride (\blacktriangle), lauroyl chloride (\bigtriangleup) and stearoyl chloride (\blacksquare).



Fig. 14. Evolution of the tensile strength (A), tensile modulus (B) and elongation at break (C) as a function of the ramie cellulose whiskers content for nanocomposite films reinforced with unmodified (\bullet) and modified with hexanoyl chloride (\circ), lauroyl chloride (\blacktriangle) and stearoyl chloride (\triangle) whiskers. The lines serve to guide the eyes.

4. Conclusions

The surface of cellulose nanocrystals or whiskers prepared by acid hydrolysis of ramie fibers was chemically modified using organic acid chloride aliphatic chains of different sizes, namely hexanoyl chloride, lauroyl chloride and stearoyl chloride. The evidence of occurrence of chemical modification was evidenced by FTIR and X-ray photoelectron spectroscopies. It was checked from X-ray diffraction analysis that the initial crystalline structure was preserved. The reduction of the polar character as determined by contact angle measurements showed that the surface chemical modification allowed enhancing the nonpolar nature of original cellulose nanocrystals, thereby allowing the use of nonpolar polymers as matrices for the processing of nanocomposite materials. Most of the studies reported in the literature with cellulose whiskers reinforced nanocomposites involve casting/evaporation processing technique. In the present study, we used a highly hydrophobic commodity plastic which is not soluble in common solvents or available in the form of latex, viz. low density polyethylene (LDPE). Indeed, almost no study of the processing of cellulose whiskers reinforced nanocomposites using industrial techniques such as extrusion has been reported. Both unmodified and functionalized nanoparticles were extruded with LDPE to prepare nanocomposite materials. The homogeneity of the ensuing nanocomposites was found to increase with the length of the grafted chains. The thermomechanical properties of processed nanocomposites were studied by DSC, DMA and tensile tests. A significant improvement in terms of elongation at break was observed when sufficiently long chains were grafted on the surface of the nanoparticles.

Acknowledgments

The authors thank CAPES (Brazil) for financial support.

References

- Bledzki AK, Gassan J. Prog Polym Sci 1999;24:221–74; Eichhorn SJ, Baillie CA, Zafeiropoulos N, Mwaikambo LY, Ansell MP, Dufresne A, et al. J Mater Sci 2001;36:2107–31;
 - Eco-composites. A special issue of. Compos Sci Technol 2003;63:114. a collection of 14 publications all dedicated to cellulose-based composite materials;
 - Mohanty AK, Misra M, Drzal LT. Natural fibers, biopolymers, and biocomposites. Boca Raton: CRC Press; 2005;
 - Jacob John M, Thomas S. Carbohydr Polym 2008;71:343-64.
- 2] Sakurada I, Nukushina Y, Ito T. J Polym Sci 1962;57:651-60.
- [3] Tashiro K, Kobayashi M. Polymer 1991;32:1516–26.
- [4] Šturcová A, Davies GR, Eichhorn SJ. Biomacromolecules 2005;6:1055-61.
- [5] Anglès MN, Dufresne A. Macromolecules 2000;33:8344–53;
 Mathew AP, Dufresne A. Biomacromolecules 2002;3:609–17;
 Orts WJ, Shey J, Imam SH, Glenn GM, Guttman ME, Revol JF, J Polym Env
- 2005:13:301-6: Kvien I. Sugiyama J, Votrubec M, Oksman K. J Mater Sci 2007;42:8163-71; Azizi Samir MAS, Alloin F, Sanchez JY, Dufresne A. Polymer 2004;45:4033-41; Azizi Samir MAS, Alloin F, Gorecki W, Sanchez JY, Dufresne A. J Phys Chem B 2004;108:10845-52; Azizi Samir MAS, Montero Mateos A, Alloin F, Sanchez JY, Dufresne A. Electrochim Acta 2004:49:4667-77 Azizi Samir MAS, Chazeau L, Alloin F, Cavaillé JY, Dufresne A, Sanchez JY. Electrochim Acta 2005:50:3897-903: Azizi Samir MAS, Alloin F, Dufresne A. Compos Interfaces 2006;13:545-59; Zimmermann T, Pöhler E, Geiger T. Adv Eng Mater 2004;6:754-61; Zimmermann T. Pöhler E. Schwaller P. Adv Eng Mater 2005:7:1156-61: Choi YJ, Simonsen J. J Nanosci Nanotechnol 2006;6:633-9. [6] Favier V, Canova GR, Cavaillé JY, Chanzy H, Dufresne A, Gauthier C. Polym Adv Technol 1995.6.351-5. Helbert W, Cavaillé JY, Dufresne A. Polym Compos 1996;17:604-11; Dufresne A, Cavaillé JY, Helbert W. Polym Compos 1997;18:198-210; Dubief D. Samain E. Dufresne A. Macromolecules 1999:32:5765-71: Dufresne A, Kellerhals MB, Witholt B. Macromolecules 1999;32:7396-401; Dufresne A. Compos Interfaces 2000;7:53-67: Chazeau L, Cavaillé JY, Canova GR, Dendievel R, Boutherin B. J Appl Polym Sci 1999.71.1797-808 Chazeau L, Cavaillé JY, Terech P. Polymer 1999;40:5333-44; Chazeau L, Paillet M, Cavaillé JY. J Polym Sci B: Polym Phys 1999;37:2151-64; Chazeau L, Cavaillé JY, Perez J. J Polym Sci B: Polym Phys 2000;38:383-92; Matos Ruiz M, Cavaillé JY, Dufresne A, Graillat C, Gérard JF. Macromol Symp 2001;169:211-22:
 - Garcia de Rodriguez NL, Thielemans W, Dufresne A. Cellulose 2006;13:261-70.

- [7] Heux L, Chauve G, Bonini C. Langmuir 2000;16:8210-2;
- Kvien I, Bjom ST, Oksman K. Biomacromolecules 2005;6:3160-5.
- [8] Goussé C, Chanzy H, Exoffier G, Soubeyrand L, Fleury E. Polymer 2002;43:2645-51;

Gopalan Nair K, Dufresne A, Gandini A, Belgacem MN. Biomacromolecules 2003;4:1835-42;

Angellier H, Molina-Boisseau S, Belgacem MN, Dufresne A. Langmuir 2005;21:2425-33.

[9] Azizi Samir MAS, Alloin F, Sanchez JY, El Kissi N, Dufresne A. Macromolecules 2004;37:386-93; Marcovich NE, Auad ML, Belessi NE, Nutt SR, Aranguren MI, I Mater Res

2006;21:870-81;

Van den Berg O, Capadona JR, Weder C. Biomacromolecules 2007;18:1353-7. [10] Nakagaito AN, Yano H. Appl Phys A 2004;78:547–52; Nakagaito AN, Iwamoto S, Yano H. Appl Phys A 2005;80:155–9;

Nakagaito AN, Yano H. Cellulose 2008;15:323–31:

Shimazaki Y, Miyazaki Y, Takezawa Y, Nogi M, Abe K, Ifuku S, et al. Biomacromolecules 2007;8:2976–8;

Yano H, Sugiyama J, Nakagaito AN, Nogi M, Matsuura T, Hikita M, et al. Adv Mater 2005:17:153-5:

Nogi M, Handa K, Nakagaito AN, Yano H. Appl Phys Lett 2005;87:243110; Iwamoto S, Abe K, Yano H. Biomacromolecules 2008;9:1022-6; Henriksson M, Berglund LA. J Appl Polym Sci 2007;106:2817-24.

- [11] Oksman K, Mathew AP, Bondeson D, Kvien I. Compos Sci Technol 2006;66:2776-84.
- [12] Bondenson D, Oksman K. Composites: Part A 2007;38:2486-92.
- Thielemans W, Belgacem MN, Dufresne A. Langmuir 2006;22:4804-10. İ13İ
- [14] Owens DK, Wendt RC. J Appl Polym Sci 1969;13:1741–7.
- [15] Habibi Y, Goffin AL, Schiltz N, Duquesne E, Dubois P, Dufresne A. J Mater Chem 2008;18:5002-10.
- [16] Freire CSR, Silvestre AJD, Pascal Neto C, Gandini A, Fardim P, Holmbom B. | Colloid Interface Sci 2006;301:205–9.
- Vaca-Garcia C, Borredon ME, Gaseta A. Cellulose 2001;8:225-31. [17]
- [18] Vaca-Garcia C, Borredon ME. Bioresour Technol 1999;70:135–42: Thiebaud S, Borredon ME. Bioresour Technol 1995;52:169-73.
- [19] Habibi Y, Dufresne A. Biomacromolecules 2008;9:1974-80.
- [20] Ferreira FC, Curvelo AAS, Mattoso LHC. J Appl Polym Sci 2003;89: 2957-65.
- [21] Azizi Samir MAS, Alloin F, Dufresne A. Biomacromolecules 2005;6: 612-26.